

High resolution integrated microbiostratigraphy of the Lower Jurassic (late Pliensbachian–early Toarcian) of central Italy

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ABSTRACT – The integrated use of calcareous nannofossil and dinoflagellate cyst events in a study of the late Pliensbachian–early Toarcian interval in central Italy has yielded a high resolution biostratigraphy. The use of both the first and last occurrences of selected taxa belonging to the two phytoplankton groups allows the dating of the sediments with a very refined detail, even when lithologies are unfavourable to the preservation of one fossil group. The evolutionary history of calcareous nannofossils and dinoflagellate cysts during the early Jurassic and its links with global events are responsible for the high potential of this integrated biostratigraphy. *J. Micropalaeontol.* 17(2): 153–172, December 1998.

INTRODUCTION

The standard biozonation for the Jurassic System is based on ammonite faunas, which possess the most important attributes for reliable biostratigraphical markers. They have restricted stratigraphical ranges, are easily identifiable, have wide geographical distributions and are generally independent of marine facies (Arkell, 1956). The considerable impetus provided by detailed worldwide geological exploration for hydrocarbon reserves (both onshore and offshore) has favoured the use of microfossils in biostratigraphy. Micropalaeontological studies have focused on Jurassic sediments, because many hydrocarbon source and reservoir rocks are of this age. Calcareous nannofossils and dinoflagellate cysts have been the object of relatively intense investigations.

Jurassic nannopalaeontology has recently received a particularly large amount of attention. Standard biostratigraphic

schemes have been proposed recently for the Jurassic (Bown, 1987; Bown *et al.*, 1988). These schemes are based mainly on the study of northern Europe, and the nannofossil events are correlated to the Boreal ammonite biostratigraphy. Some problems with the application of previous schemes to lower latitudes arise because of provincialism both of nannoplankton and ammonites. Research in the Tethyan region has proceeded at a lower pace, due mainly to the presence, at various levels, of unfavourable lithologies. Recently, a synthesis of calcareous nannofossil events in the Tethyan Realm has been attempted by Mattioli & Erba (in press). The interval in which good biostratigraphic resolution is provided by nannofossils is the Pliensbachian–Bathonian, and in particular in the late Pliensbachian–early Bajocian, where nannofossils display a biostratigraphic potential comparable to that of the ammonites.

The first dinoflagellate cyst biozonation for the entire Jurassic

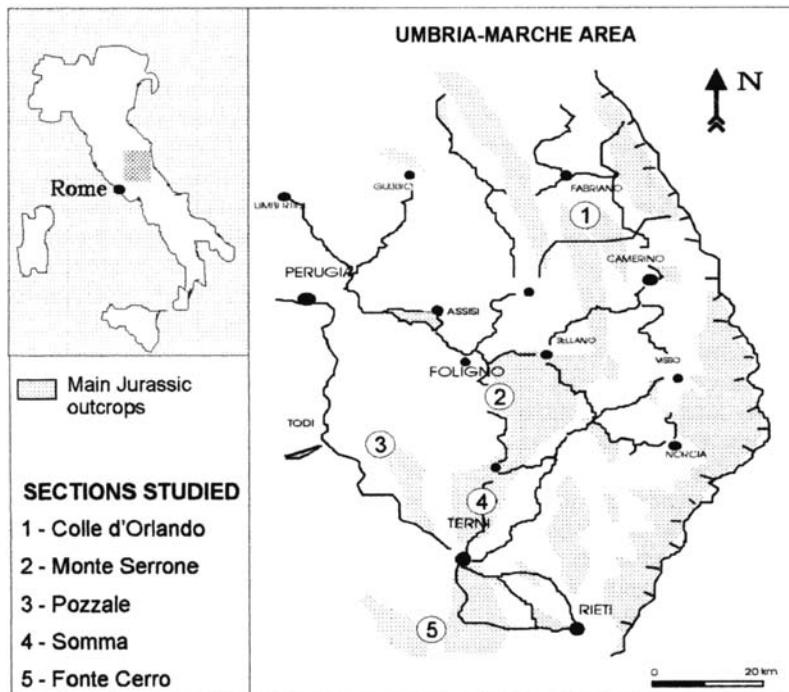


Fig. 1. Location map of the sections studied.

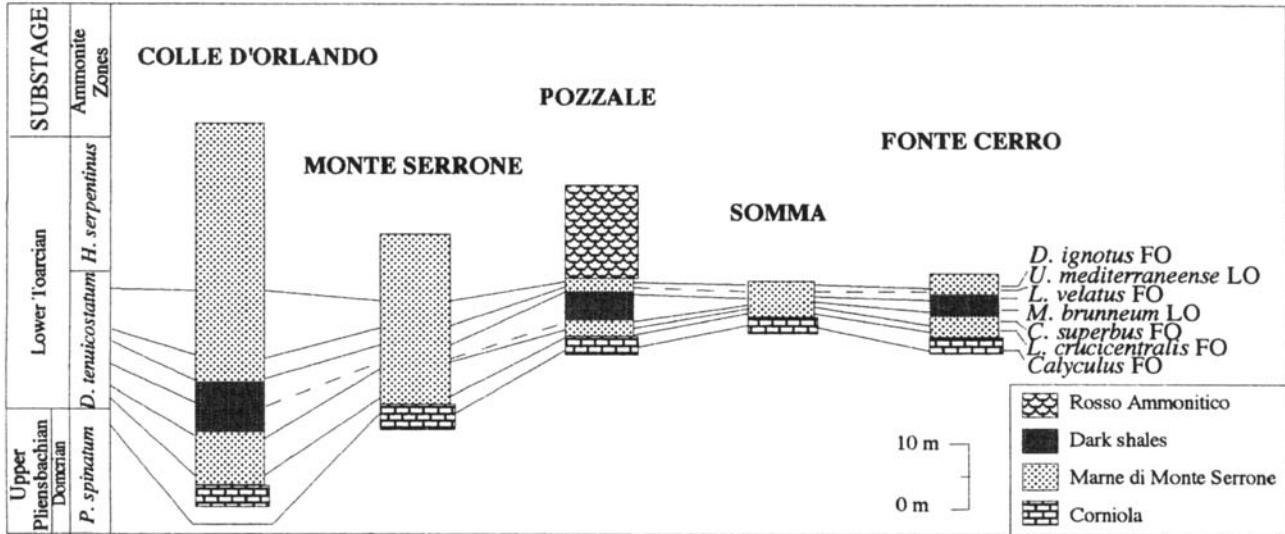


Fig. 2. Lithology and biostratigraphy of the sections studied. Correlation lines based on the most significant calcareous nannofossil and dinoflagellate cyst events are illustrated (FO = first occurrence; LO = last occurrence). Where a bioevent has not been recorded a dashed line has been used.

System was proposed by Woollam & Riding (1983) from the study of boreholes and outcrop samples from England. In this biozonation, the boundaries of the dinoflagellate cyst zones were compared to those of the ammonites. More recently, Riding & Thomas (1992) revised the biozonation of Woollam & Riding (1983) including data from southern and eastern England and western Scotland. Dinoflagellate cysts exhibit lower resolution than ammonites in the Early and Mid-Jurassic, and have approximately the same biostratigraphic resolution as ammonites during the Late Jurassic.

The presence of rich and well-preserved assemblages of calcareous nannofossils and dinoflagellate cysts depends on several factors, such as lithology, palaeoenvironmental conditions and diagenetic overprint. Organic matter is preferentially concentrated in fine grained sediments deposited under dysaerobic-anoxic conditions (Tyson, 1987). It is lacking in coarse grained and calcareous sediments which were deposited in highly oxidizing regimes in which the organic constituents are rapidly degraded. Calcareous nannofossil preservation is strongly dependent on the lysocline depth in the seas, burial conditions, and the presence and diagenesis of organic matter. Well preserved assemblages are generally observed in marly lithotypes rather than in argillaceous and calcareous ones, where post-burial conditions often severely alter the assemblage composition (Mattioli, 1995). Frequently, the two phytoplankton fossil groups show their highest concentration and better preservation in different lithologies: calcareous nannofossils are common in limestones and marly limestones, while dinoflagellate cysts are more abundant in bituminous marls and shales.

Ecological conditions, such as trophic resources, may profoundly affect the composition of phytoplankton assemblages. According to Hallock (1987) coccolithophorids may grow in low concentrations of nutrients, whereas dinoflagellates generally dominate in mesotrophic conditions. The different ecology and lithological preferences for their preservation

suggest that their integration may provide a detailed biostratigraphical framework, independent of sedimentary facies.

Umbria-Marche (central Italy) is an important biostratigraphic reference area for the Tethyan Realm, because of the continuity of numerous successions and the rich micro- and macrofossil content. Moreover, it represents a key region in which to understand the evolution of the western Tethys during the Jurassic. Due to the presence of several sections which are well dated by ammonites, Umbria-Marche is particularly suitable for carrying out integrated biostratigraphy based on calcareous nannofossils and dinoflagellate cysts. This study focuses on the late Pliensbachian-early Toarcian, an interval characterized by numerous global palaeoecological and palaeogeographical events (i.e. speciations, transgression and anoxia).

The aims of this paper are:

- the description of calcareous nannofossil and dinoflagellate cyst distributions in the Early Jurassic of central Italy;
- the identification of the most significant bioevents in the two phytoplankton groups;
- a bioevent integration in order to obtain a high resolution biostratigraphic scheme.

GEOLOGICAL SETTING

The Umbria-Marche area is located in the southerly part of the Northern Apennines, delimited by the Sillaro Valley to the north and the Latium-Abbruzzi carbonate platform to the south (Fig. 1). The Jurassic successions were deposited in a basin which originated during a phase of extensional tectonics, related to the opening of the western Tethys and inducing the breakup and drowning of the Calcare Massiccio carbonate platform (Colacicchi *et al.*, 1989; Cresta *et al.*, 1989). The palaeotopography was quite complex as a consequence of rifting and subsequent differential subsidence: structural highs were connected by more or less steep slopes to the troughs. This palaeogeographical framework is reflected in the different depositional patterns

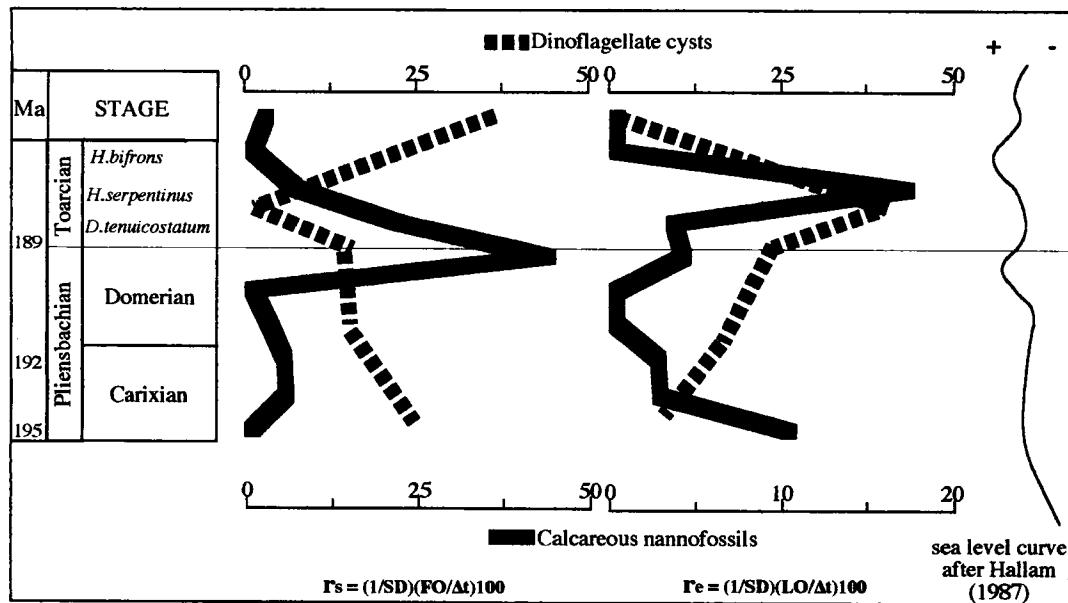


Fig. 3. Evolutionary rates of the calcareous nannofossils and dinoflagellate cysts related to the eustatic curve of Hallam (1987). The speciation and extinction rates have been calculated according to Roth (1986). The time scale used is from Gradstein *et al.* (1994). The number of phytoplankton species has been obtained from integration of the published data with the present ones.

which occurred. On the structural highs, condensed sequences can be recognized, whereas in the deepest depressions thick and continuous sequences with several resedimented levels were deposited. The most common situation is represented by continuous sequences with intermediate thicknesses and without frequent resedimented levels. The subsidence and relative sea-level reached their maximum during the Toarcian, when argillaceous lithotypes were deposited throughout the basin. Anoxic conditions developed in most of the western Tethys at that time, probably as consequence of the sea level rise (Hallam, 1987; Jenkyns, 1985).

MATERIAL AND METHODS

Five successions were sampled in the Umbria-Marche area, in order to document all the different sectors of the basin, following a transect from the areas close to the Latium-Abruzzi carbonate platform and influenced by its evolution to the most distal areas (Fig. 2).

The Colle d'Orlando section is located in the northeastern area of the Umbria-Marche Basin (Fig. 1). The examined interval is early Toarcian in age, between two horizons containing several specimens of *Dactylioceras* spp. In central Umbria, the Monte Serrone and Somma sections have been investigated. The interval studied in the Monte Serrone section comprises the late Domerian, *P. spinatum* Zone, the early Toarcian *D. tenuicostatum* Zone and *H. serpentinus* Zone (Reale *et al.*, 1992). In the Somma section, because of the lack of ammonites, the biostratigraphy is based on calcareous nannofossils and dinoflagellate cysts. The Pozzale section is in central western Umbria and it is dated as late Domerian-early Toarcian, according to the ammonite faunas (Mattioli, 1995; Nini *et al.*, 1995). The Fonte Cerro section is located in the Monti Sabini area (north Latium). Some ammonite horizons give an age of *D. tenuicostatum* to *H. serpentinus* Zones for the

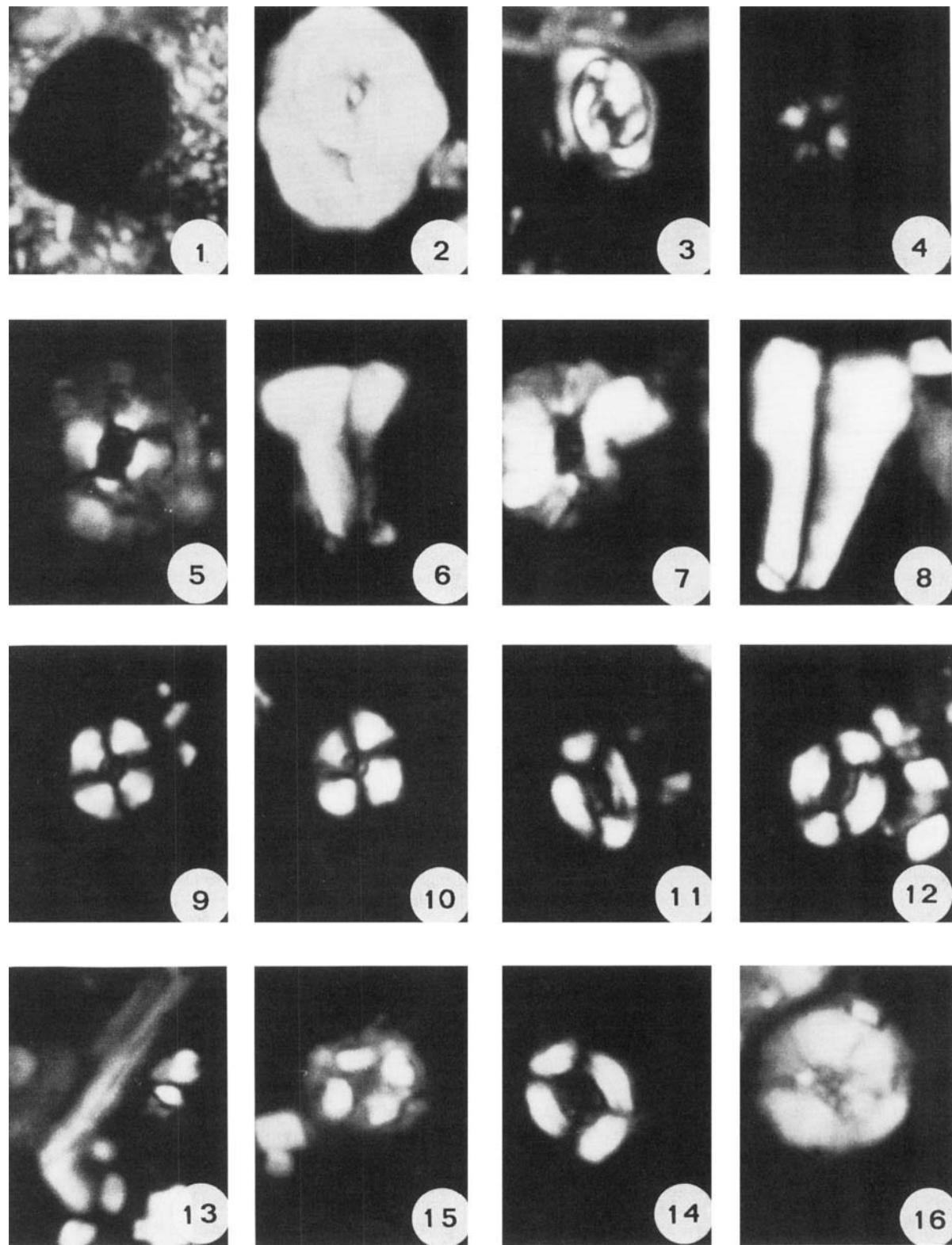
studied interval (Bucefalo Palliani & Mattioli, 1994).

The lithotypes studied comprise: (1) the uppermost levels of the Corniola Unit, nutty to white micritic limestones with occasional detrital levels interbedded in some cases with chert in layers or nodules, Domerian in age; (2) the Marne di Monte Serrone Formation (Pialli, 1969), marls and argillaceous marls yellow to dark in colour, with occasional calcarenites, which may include black shale levels, early Toarcian in age; (3) the basal portion of the Rosso Ammonitico Unit, nodular marls and limestones, red to greenish, in which the clay content decreases upwards, early to mid-Toarcian in age.

Samples were generally taken at regular intervals, from more micritic limestones and/or from marlstones, trying to avoid the detrital levels. Sample spacing varied from 5 to 20 cm. Quantitative studies on calcareous nannofossil and dinoflagellate cyst assemblages have been performed on the same samples. Samples for observations of calcareous nannofossils under a light microscope were prepared with standard techniques, described by Mattioli (1995). Palynological slides were obtained by standard preparation procedures, involving acid treatment, oxidative maceration and sieving using a 10 µm mesh (Wood *et al.*, 1996).

CALCAREOUS NANNOFOSSIL ASSEMBLAGES

A total of 313 samples were quantitatively studied. Calcareous nannofossil preservation varies from poor to moderate depending on the different lithotypes. The poorest preservation was observed in the Colle d'Orlando and Fonte Cerro sections and, concomitantly with (a) detrital limestones, in which the high primary porosity allowed the aggressive diagenetic waters to circulate, and (b) in the argillaceous layers in which the highest values of Total Organic Carbon (TOC) have been recorded, in relation to the diagenesis of the organic matter. The predominant lithotypes are represented by marly limestones, ensuring a



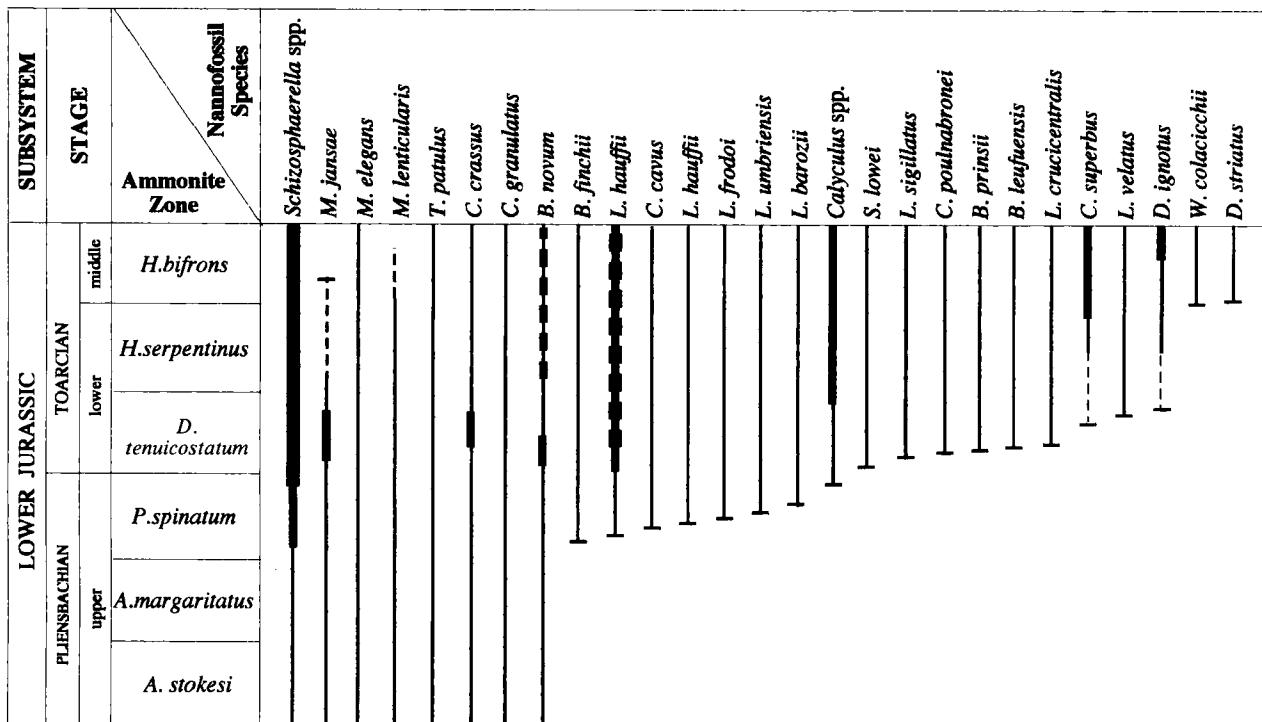


Fig. 4. Stratigraphical distribution of calcareous nannofossils from the Lower Jurassic of central Italy. The line thickness is related to the relative abundances of each species in the assemblages. Dashed lines indicate discontinuous occurrences.

moderately good nannofossil preservation, and the sample density is high. The effects of diagenetic impoverishment can be considered negligible for the following biostratigraphic considerations.

All the sections studied display comparable nannofossil abundance and preservation, although the Pozzale and Monte Serrone successions seem to be the richest (Appendix B, Figs B1–B6). The data presented in this paper are supported by the dataset collected in several other successions studied in central Italy (Mattioli, 1995).

A significant change was observed across the Domerian–Toarcian boundary in the assemblage composition of calcareous nannofossils, as at that time the diversification rate is the highest of the entire Jurassic System (Bown, 1996) and the speciations largely dominate over the extinctions (Fig. 3). The range charts of all the sections studied are illustrated in Appendix B (Figs B1–B6). The biostratigraphical data are summarized in Fig. 4.

In the latest Domerian (top of the *P. spinatum* Zone) the

assemblages are low in diversity and never abundant. The prominent taxa in the assemblages are represented by the *incertae sedis* *Schizosphaerella* spp., *Mitrolithus jansae*, *Crepidolithus crassus*, rare specimens of the genus *Biscutum* (i.e. *B. dubium*) and genus *Lotharingius* (such as *L. hauffii* and *L. barozii*). After a long period of evolutionary quiescence which lasted the entire Pliensbachian, several new genera and species first appeared. Among these, some had a successful evolutionary history, being long ranging species. The first appearances of *Crepidolithus cavus* (= *C. impontus* of some authors), *Biscutum finchii* and *Biscutum grande* are significant. These species have a distribution restricted to the Tethyan Realm (Bown, 1987; Mattioli & Erba, in press). A great diversification in the Family Watznaueriaceae is observed in this interval as many different species appeared, such as *Lotharingius hauffii*, *L. barozii* and *L. sigillatus*. *Lotharingius umbriensis* and *L. frodoi* have been recently described by Mattioli (1996) from the Lower Toarcian of western Tethys. The first occurrences of *L. hauffii* and *L.*

Explanation of Plate 1

All light micrographs, crossed nicols, approximately $\times 4600$. **fig. 1.** *Schizosphaerella* sp. indet. Deflandre & Dangeard, 1938. Sample PO 7.20, Pozzale section. **fig. 2.** *Crepidolithus crassus* (Deflandre, 1954) Noël, 1965. Sample PO 7.20, Pozzale section. **fig. 3.** *Tubirhabdus patulus* Rood *et al.*, 1973. Sample MS1 19.20, Monte Serrone section. **fig. 4.** *Biscutum dubium* (Noël, 1965) Grün in Grün *et al.*, 1974. Sample PO 7.20, Pozzale section. **fig. 5.** *Biscutum grande* Bown, 1987. Sample PO 7.20, Pozzale section. **fig. 6.** *Calyculus* sp. indet.. Sample MS1 19.20, Monte Serrone section. **fig. 7.** *Calyculus* sp. indet.. Sample MS1 19.20, Monte Serrone section. **fig. 8.** *Carinolithus poulnabronei* Mattioli, 1996. Sample MS1 19.20, Monte Serrone section. **fig. 9.** *Lotharingius hauffii* Grün & Zweili in Grün *et al.*, 1974. Sample MS1 19.20, Monte Serrone section. **fig. 10.** *Lotharingius frodoi* Mattioli, 1996. Sample MS1 19.20, Monte Serrone section. **fig. 11.** *Bussonius prisii* (Noël, 1973) Goy, 1979. Sample MS1 19.20, Monte Serrone section. **fig. 12.** *Lotharingius crucicentralis* (Medd, 1971) Grün & Zweili, 1980. MS1 19.20, Monte Serrone section. **fig. 13.** *Carinolithus superbus* (Deflandre, 1954) Prins in Grün *et al.*, 1974. Sample MS1 19.20, Monte Serrone section. **fig. 14.** *Lotharingius velatus* Bown & Cooper, 1989. Sample MS1 19.20, Monte Serrone section. **fig. 15.** *Discorhabdus ignotus* (Gorka, 1957) Perch-Nielsen, 1968. Sample PO 7.20, Pozzale section. **fig. 16.** *Discorhabdus striatus* Moshkovitz & Ehrlich, 1976. Sample PO 7.20, Pozzale section.

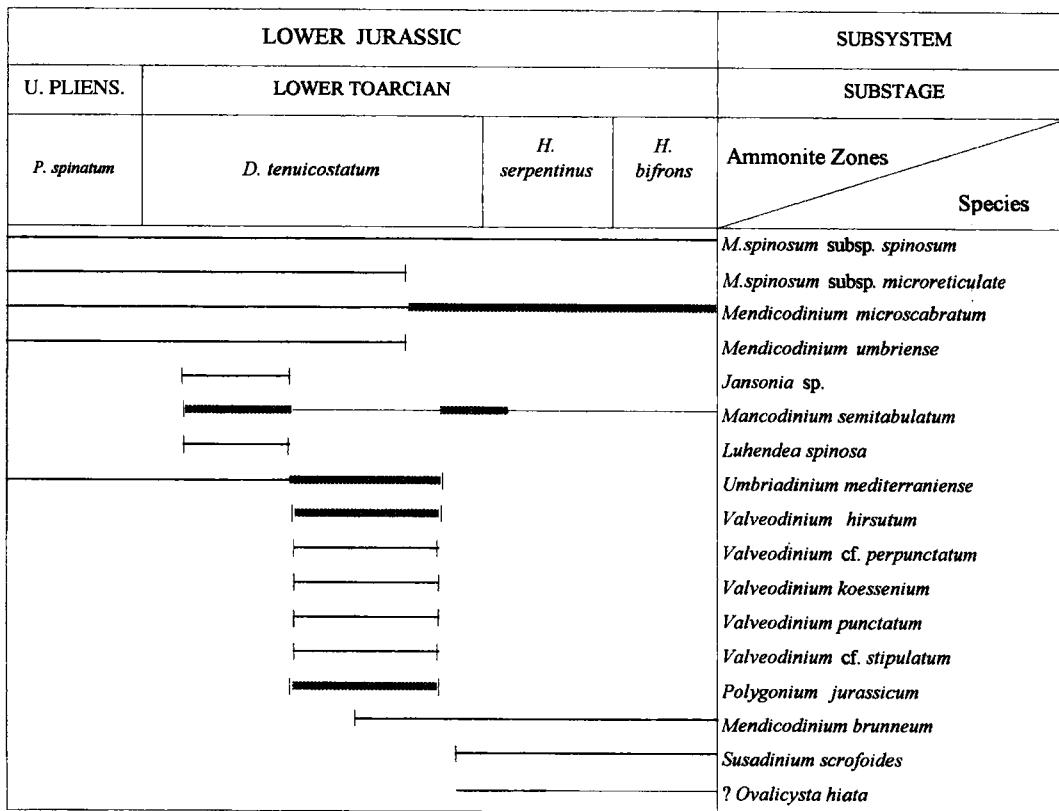


Fig. 5. Stratigraphical distribution of dinoflagellate cysts from the Lower Jurassic of central Italy. The line thickness is related to the relative abundances of each species in the assemblages.

sigillatus seem to be two reliable events recorded concordantly in both the Tethyan and Boreal domains. The entry of the Family Calyculaceae with the genus *Calculus* is also observed. This event constitutes a significant marker for the Pliensbachian–Toarcian boundary in the Tethyan Realm.

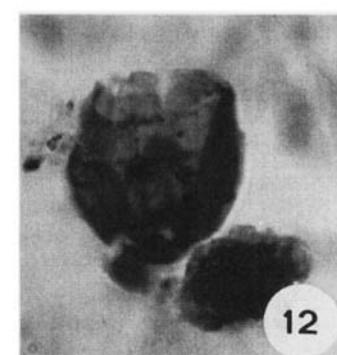
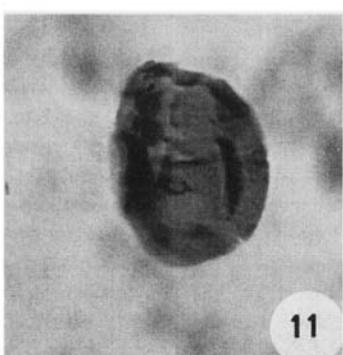
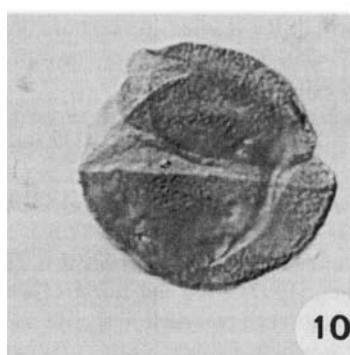
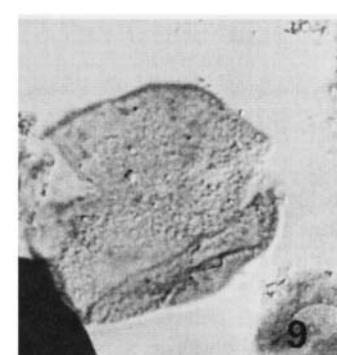
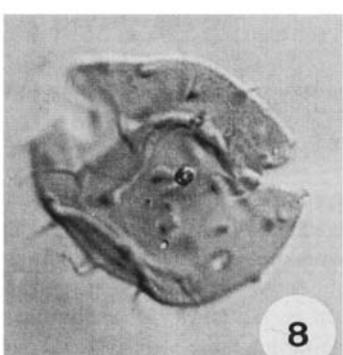
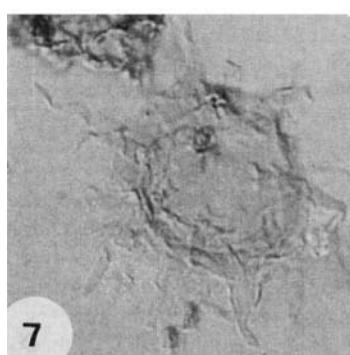
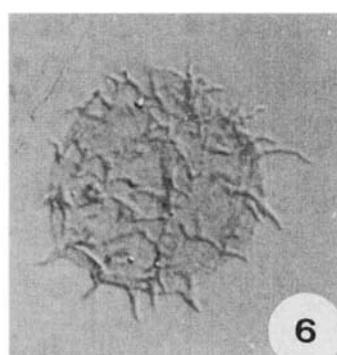
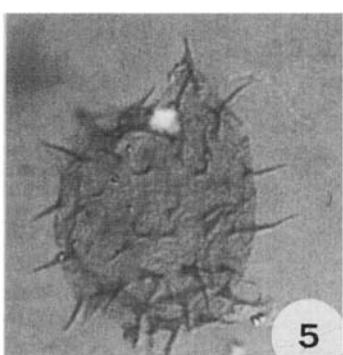
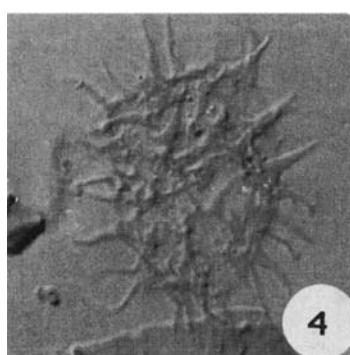
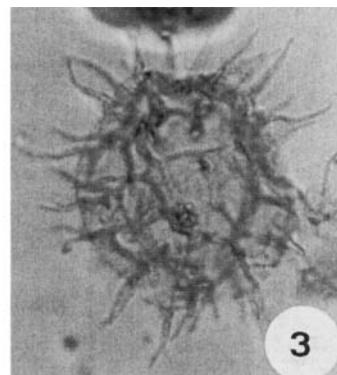
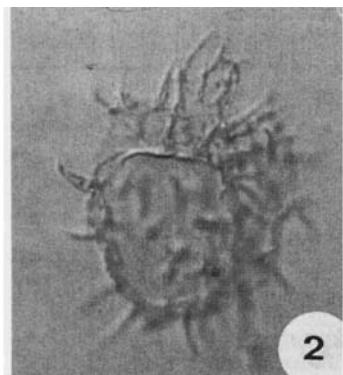
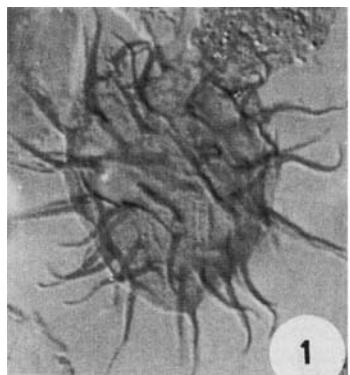
This diversification trend continued into the early Toarcian *D. tenuicostatum* Zone, in which the genus *Carinolithus* appeared with *C. poulnabronei* and *C. superbus*, which are however rare immediately after their appearance, becoming common in the mid-Toarcian. Among the Watznaueriaceae, rare specimens of *Bussonius prinsii* and *B. lefuensis*, *Lotharingius crucicentralis* and *L. velatus* first appear, which, although rare, are typical constituents of Toarcian assemblages. In the uppermost part of this zone the easily recognizable *Discorhabdus ignotus* has its first appearance. Also *D. ignotus* is very rare after its appearance but

becomes abundant in the mid-Toarcian, probably due to favourable palaeoenvironmental conditions. In the early Toarcian *H. serpentinus* Zone, the important genus *Watznaueria*, which dominates Mid and Late Jurassic assemblages, appears with *W. colacicchii* and *Watznaueria* sp. 1. The significant taxon *Mitrolithus jansae*, which dominated the early Jurassic assemblages, abruptly decreases in abundance in advance of its disappearance.

The assemblages are not rich in the Domerian and lowermost part of the Toarcian, both in species diversity and total abundance. In the early *D. tenuicostatum* Zone, a notable increase of species diversity and total abundance occurred. Although many variations are distinguished in the assemblage composition, and in the late *D. tenuicostatum* Zone the nannoplankton experienced a crisis, the early Toarcian is

Explanation of Plate 2

All photomicrographs taken in plain transmitted light, magnification $\times 1250$. **fig. 1.** *Valveodinium hirsutum* Bucefalo Palliani & Riding, 1997. Sample CO 14.80 (2), England Finder coordinate Q 36, Colle d'Orlando section. **fig. 2.** *Valveodinium hirsutum* Bucefalo Palliani & Riding, 1997. Sample CO 8.50, England Finder coordinate G 42-1/3, Colle d'Orlando section. **fig. 3.** *Umbriadinium mediterraneense* Bucefalo Palliani & Riding, 1997. Sample FC1 0.70 (6), England Finder coordinate Q 47-4, Fonte Cerro section. **fig. 4.** *Umbriadinium mediterraneense* Bucefalo Palliani & Riding, 1997. Sample FC1 0.70 (6), England Finder coordinate L 35, Fonte Cerro section. **fig. 5.** *Umbriadinium mediterraneense* Bucefalo Palliani & Riding, 1997. Sample CO 8.50, England Finder coordinate S 37, Colle d'Orlando section. **fig. 6.** *Umbriadinium mediterraneense* Bucefalo Palliani & Riding, 1997. Sample PO 3.60 (2), England Finder coordinate L 38, Pozzale section. **fig. 7.** *Polygonium jurassicum* Bucefalo Palliani et al., 1996. Sample FC1 1.00 (1), England Finder coordinate K 28, Fonte Cerro section. **fig. 8.** *Mendicodinium spinosum* subsp. *perforatum* Bucefalo Palliani et al., 1997. Sample PO 3.60 (4), England Finder coordinate H 24-3, Pozzale section. **fig. 9.** *Mendicodinium microscabratum* Bucefalo Palliani et al., 1997. Sample CO 6.90, England Finder coordinate R 36, Colle d'Orlando section. **fig. 10.** *Mendicodinium brunneum* Bucefalo Palliani et al., 1997. Sample CO 13.10 (1), England Finder coordinate D 31-3, Colle d'Orlando section. **fig. 11.** *Jansonia* sp. Sample CO 7.30 (1), England Finder coordinate W 34, Colle d'Orlando section. **fig. 12.** *Jansonia* sp. Sample CO 6.90 (2), England Finder coordinate O 39-3, Colle d'Orlando section.



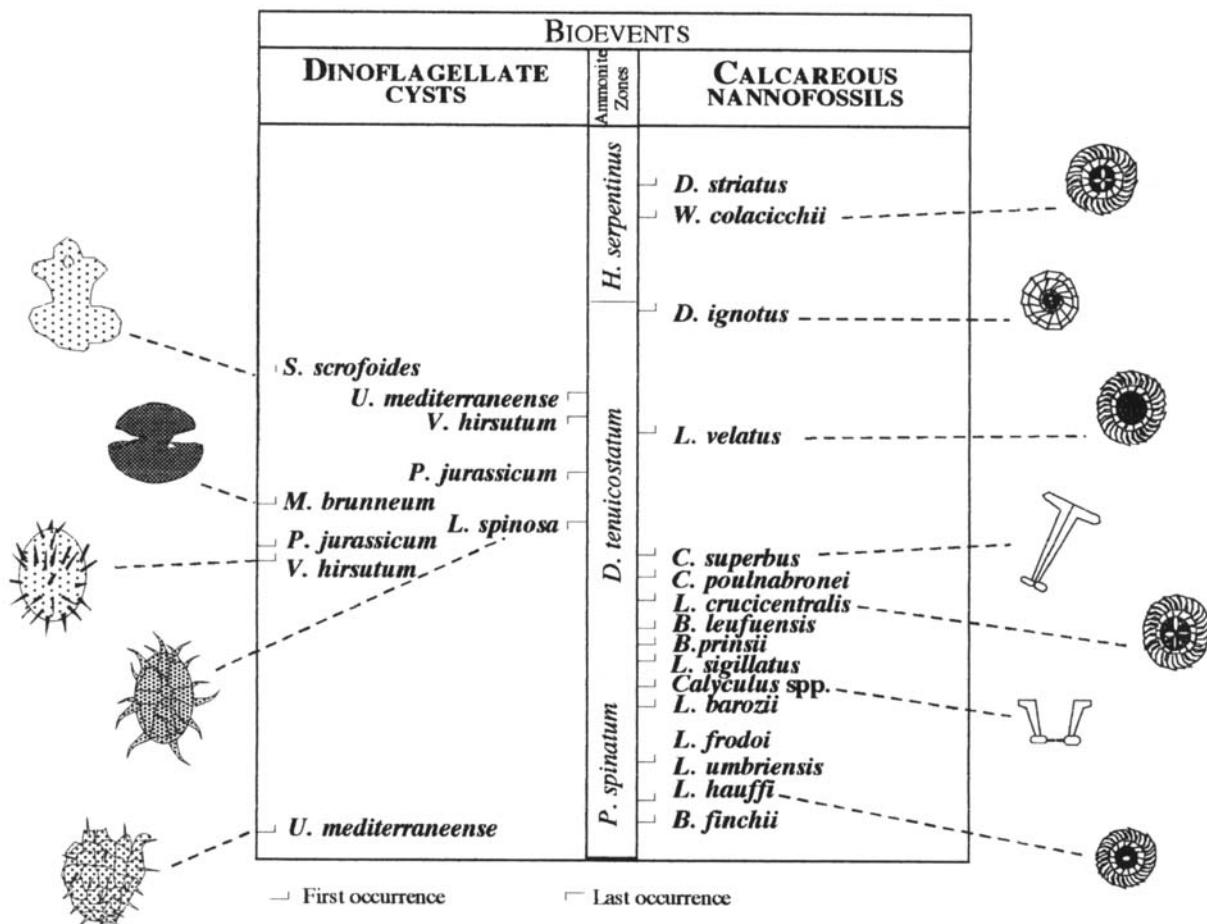


Fig. 6. Integrated phytoplanktonic events, both first and last occurrences, with regards to the ammonite zones during the late Pliensbachian–early Toarcian.

generally an interval in which great abundances and a diversified assemblage are observed in the areas studied. Large fluctuations in the abundance of each taxon, depending on palaeoecological conditions, were observed by Bucefalo Palliani & Mattioli (1995). Although palaeoecology and preservation often induced fluctuations of abundance, a certain continuity is observed in the occurrences of various species. The continuity in the findings is also a consequence of the close spaced sampling, which allowed the recognition of the original distribution of nannofossils.

DINOFLAGELLATE CYST ASSEMBLAGES

Forty samples were studied for palynomorphs. In central Italy the Pliensbachian lithologies comprise calcareous lithotypes and are not favourable to palynological analyses. The rare marly intercalations generally are palynologically barren. The only information regarding the late Pliensbachian (*P. spinatum* Zone) is from the Fonte Cerro section (Appendix C, Fig. C5). The lower Toarcian lithologies (Marne di Monte Serrone Formation) are promising for palynological investigations. Their organic content and the variations of the terrestrial and marine organic constituents are high and related to eustatic fluctuations (Bucefalo Palliani & Cirilli, 1993; Bucefalo Palliani *et al.*, in

press). The most abundant and diverse dinoflagellate cyst assemblages have been recorded in the sections characterized by bituminous lithologies within the Marne di Monte Serrone Formation (i.e. Colle d'Orlando, Pozzale and Fonte Cerro). In other sections, the dinoflagellate cyst data are rare (i.e. Somma and Monte Serrone). However, the integration of the data from all the sections studied, allows the elucidation of the dinoflagellate cyst distribution in the Lower Toarcian of central Italy. The range charts of all the studied sections are illustrated in Appendix C (Figs C1–C5). The biostratigraphical data have been summarized and illustrated in Fig. 5.

In the late Pliensbachian (Domerian) and early Toarcian (early *D. tenuicostatum* zone) the dinoflagellate cyst assemblages are characterized by rare specimens of *Mendicodinium* (*M. spinosum* subsp. *spinosum*, *M. spinosum* subsp. *perforatum*, *M. microscabratum* and *M. umbriense*) and by *Umbriadinium mediterraneense*. The *Mendicodinium* species were first described by Bucefalo Palliani *et al.* (1997) from the Lower Toarcian of central Italy. They have also been recorded in the Pliensbachian and Lower Toarcian of southern France, Greece, Hungary and Portugal (Baldaña *et al.*, 1995; Bucefalo Palliani, 1996; Bucefalo Palliani & Riding, 1998). According to Bucefalo Palliani (1996) the base of the Pliensbachian in the Tethyan

domain is marked by the first occurrence of the '*Mendicodinium* group', consisting of small cysts with varying wall ornamentation. The several species of *Mendicodinium* are rarely present throughout the Lower Toarcian (*D. tenuicostatum* Zone). *Mendicodinium microscabratum* has been recorded up to the late Toarcian in southern France (Bucefalo Palliani & Riding, 1998). *Umbriadinium mediterraneense* is a suessiacean dinoflagellate cyst described by Bucefalo Palliani & Riding (1997a) from northwest Greece, central Italy and Hungary. In central Italy its stratigraphical range is from late Pliensbachian (*P. spinatum* Zone) to early Toarcian (*D. tenuicostatum* Zone). It is rare in the late Pliensbachian and lower part of the *D. tenuicostatum* Zone and becomes abundant in the middle–upper part of the *D. tenuicostatum* Zone. The range top of *U. mediterraneense* in central Italy is at the top of the *D. tenuicostatum* Zone.

Mancodinium semitabulatum has been recorded from the entire *D. tenuicostatum* Zone. It is a long ranging (Pliensbachian–Bajocian) and cosmopolitan taxon, first described from the Upper Pliensbachian of Germany (Morgenroth, 1970). In central Italy, *M. semitabulatum* exhibits a decrease in abundance in the middle portion of the *D. tenuicostatum* Zone, within the most bituminous lithotypes. This trend has been interpreted as the result of the dinoflagellate life strategies and the stable palaeoecological conditions of the palaeoenvironment (Bucefalo Palliani, 1996). Rare specimens of *Luehndea spinosa* have been recorded in the Fonte Cerro and Pozzale sections (Appendix C, Figs C3 & C5) in the lower part of the *D. tenuicostatum* Zone. The range of *L. spinosa* is restricted to the late Pliensbachian–earliest Toarcian (Riding, 1987) in northern Europe. In the lower portion of the *D. tenuicostatum* Zone of the Colle d'Orlando section, rare specimens of *Jansonia* sp. have been reported. *Jansonia* sp. exhibits the same wall ornamentation of the Bathonian *Jansonia manifesta* but with lower parasutural ridges. The parasutural ornamentation of *Jansonia* sp. is similar to that of *Jansonia jurassica*. The low number of specimens recorded prevents a detailed and formal description of this species.

In the middle part of the *D. tenuicostatum* Zone the dinoflagellate cyst assemblages from central Italy record the highest species diversity (Fig. 5). The assemblages are characterized by several species of *Valvaeodinium*, such as *V. hirsutum*, *V. koessenum*, *Valvaeodinium* cf. *perpunctatum*, *V. punctatum* and *V. stipulatum*. *Valvaeodinium punctatum*, *V. perpunctatum* and *V. stipulatum* were first described as *Comparodinium* from the late Pliensbachian–early Toarcian of southwest Germany (Wille & Gocht, 1979). *Valvaeodinium koessenum* was recorded by Morbey (1975) from the Rhaetian of Austria. In the Pliensbachian and Lower Toarcian, Wille & Gocht (1979) recognized *Comparodinium* cf. *koessenum*, characterized by the lack of superficial microreticulation. *Valvaeodinium hirsutum* was described by Bucefalo Palliani & Riding (1997a) from the Lower Toarcian of Greece and central Italy.

In the middle portion of the *D. tenuicostatum* Zone, abundant specimens of the acritarch *Polygonium jurassicum* Bucefalo Palliani *et al.* 1996 and the first occurrence of *Mendicodinium brunneum* have also been recorded. This species of *Mendicodinium* has been reported in the Lower Toarcian (*D. tenuicostatum* Zone) of Greece, the Lower Toarcian (*D. tenuicostatum* Zone)

and *H. serpentinus* Zone) of Portugal and the Upper Toarcian (*Grammoceras thouarsense* Zone) of southern France (Bucefalo Palliani, 1996; Bucefalo Palliani & Riding, 1998).

In the upper part of the *D. tenuicostatum* Zone, *Susadinium scrooides* and *Ovalicysta? hiata*, taxa belonging to the *Parvocysta* suite of Riding (1984), first occurred. These bioevents have been recognized by Bucefalo Palliani & Riding (1997b) and Bucefalo Palliani & Mattioli (1994). The discrepancies regarding the first occurrence of the *Parvocysta* suite in the Tethyan and Boreal domains have been explained through a mid-Toarcian migrational event, related to the gradual oxygenation of the Boreal seas following the early Toarcian anoxic event (Bucefalo Palliani & Riding, in press).

The palynological data from the *H. serpentinus* Zone reveal low diversity dinoflagellate cyst assemblages, consisting of *M. semitabulatum*, *M. microscabratum*, *M. brunneum* and representatives of the *Parvocysta* suite.

INTEGRATED BIOSTRATIGRAPHY

The large number of phytoplankton events recorded in the late Pliensbachian and early Toarcian makes possible the selection of a certain number of first and last occurrences (FO and LO, respectively) of species which are useful for biostratigraphical correlation. Only species with a well defined stratigraphical range have been considered. The occurrences of these species are calibrated with ammonites and recorded in central Italy as in other Tethyan and sometimes Boreal domains. Where possible, the taxa which are more resistant to diagenetic processes have been chosen. In some cases more delicate taxa have been considered, such as the calcareous nannofossil *D. ignotus* and the dinoflagellate cyst *U. mediterraneense*, because of their stratigraphical importance and easily recognizable morphological characteristics. Due to a careful taxonomic revision and the introduction of a new genus and some new species, all the species exhibit a stable and well defined taxonomy (Bucefalo Palliani *et al.*, 1996; Bucefalo Palliani & Riding, 1997a; Mattioli, 1996). The selected species display characteristic and easily recognizable features under the light microscope, and are widespread and commonly found throughout the Tethyan domain.

From the integrated scheme of Fig. 6, an interplay between the two fossil groups is evident. A more detailed biostratigraphical resolution for the Pliensbachian–Toarcian boundary is provided by calcareous nannofossils, whereas the dinoflagellate cysts yield greater detail in the uppermost part of the *D. tenuicostatum* Zone. The nannofossil resolution is again high between the *D. tenuicostatum* and *H. serpentinus* zones. The great detail provided by phytoplankton in this time interval is linked to their evolutionary history, characterized by speciation events and by different diversification and turnover trends. The high potential of the calcareous nannofossil and dinoflagellate cyst integration lies in the slight diachroneity of the speciation and turnover events recorded by the two phytoplanktonic groups (Fig. 3). The notable speciation event experienced by calcareous nannofossils at the transition between the Pliensbachian and Toarcian is concomitant with the inception of the early Toarcian transgression, which increased the number of available niches. The dinoflagellate cyst record indicates high

turnover, concomitant with the stressed palaeoenvironmental conditions during the early Toarcian anoxic event (Fig. 3). The low oxygen content at the sea bottom diminished the number of available niches for the cyst-producing dinoflagellates.

The first event recorded in the late Pliensbachian (*P. spinatum* Zone) is the FO of *U. mediterraneense*. An older occurrence of this taxon is not excluded, because the poor preservational potential of the late Pliensbachian lithologies prevents the collection of detailed palynological data. The Pliensbachian–Toarcian boundary is marked by a succession of 12 nannofossil events (Fig. 6). Several species of *Lotharingius* appear, suggesting an acme in the evolutionary history of this genus. Among these events the FOs of *L. hauffii* (*P. spinatum* Zone) and *L. crucicentralis* (*D. tenuicostatum* Zone) are significant, because of their good recovery in several different areas. The Pliensbachian–Toarcian boundary is also defined by the entry of the Genus *Calyculus*. This event has always been referred to as Late Domerian in the Tethyan Realm (for a revision, see Mattioli & Erba, in press), although in North Europe it is reported in the early Domerian (Crux, 1984) or as sporadic from the Mid-Carixian and continuous from the Late Domerian (Bown, 1987; Bown *et al.*, 1988). In the basal Toarcian two species of the genus *Bussinius* first appeared. As these specimens are delicate, an older occurrence in the late Pliensbachian limestones of the Corniola Unit is not excluded. Another important genus appearing within the *D. tenuicostatum* Zone is *Carinolithus*, with the two subsequent easily recognizable species *C. poulnabronei* and *C. superbus*. Between the FOs of *C. superbus* and *L. velatus*, several dinoflagellate cysts occur. The FO of *V. hirsutum* is synchronous with the FO of *C. superbus* which is followed by the FO of the acritarch *P. jurassicum* and by the LO of *L. spinosa*. *Polygonium jurassicum* has a very restricted range and it disappears within the *D. tenuicostatum* Zone. The LO of *L. spinosa*, a well calibrated and recognizable event both in the Boreal and Tethyan realms, is followed by the FO of *M. brunneum*. Above the FO of *L. velatus*, the subsequent LO of *V. hirsutum* and *U. mediterraneense* is verified. The last dinoflagellate event in the *D. tenuicostatum* Zone is the FO of *S. scrofoides*, which represents the first species of the Heterocapsaceae lineage. The end of the *D. tenuicostatum* Zone corresponds to the entry of the genus *Discorhabdus* with the species *D. ignotus*. No dinoflagellate events are recorded in the *H. serpentinus* Zone and this may reflect a period of relative evolutionary quiescence. Nannoplankton diversification increased again in the late *H. serpentinus* Zone, with the FO of *D. striatus* and the entry of the important genus *Watznaueria*, which dominates all later Jurassic assemblages.

DISCUSSION AND CONCLUSIONS

Integration of selected calcareous nannofossil and dinoflagellate cyst events has produced a detailed biostratigraphical framework for the late Pliensbachian–early Toarcian of central Italy. This scheme demonstrates that the integration of phytoplankton data may represent an important parachronology to the ammonite zonation, which provides the orthochronology of the Jurassic. The integration of different phytoplankton groups ensures more precise biochronology and the ability to date sediments even when lithologies unfavourable to the preserva-

tion of one fossil group are present.

The success of this biostratigraphical approach derives from the different evolutionary history of the two phytoplankton groups. The slight diachronity between the early Jurassic radiation of nannoplankton and dinoflagellate cysts is linked to the interplay of phytoplankton life cycles and global events. The calcareous nannoplankton exhibit a maximum in diversification at the Pliensbachian–Toarcian transition, probably produced by the inception of early Toarcian sea level rise. It is coincident with the negative excursion of the $\delta^{13}\text{C}$ curve of Jenkyns & Clayton (1997), about 1 Ma before the oceanic anoxic event (Fig. 3) and the positive excursion of the $\delta^{13}\text{C}$ curve of Jenkyns & Clayton (1997). The early Jurassic sea level rise produced high diversification rates within dinoflagellate community, earlier than for nannoplankton, in the early Pliensbachian (Mattioli & Bucefalo Palliani, 1995). Concomitant with the stressed conditions, related to the inception of the global anoxic event, dinoflagellates exhibit high turnover, while nannoplankton had a low diversification period. After the global anoxic event a second diversification pulse of nannoplankton occurred, which saw the appearance of the important *Discorhabdus* and *Watznaueria* genera. With regard to dinoflagellates, the following diversification pulse is in the *H. bifrons* Zone, when the recovery of the oxygenated conditions at the sea bottom produced an increase of the available habitats.

The differences between the evolutionary trends of the two phytoplanktonic groups can be ascribed to their life cycles. Nannoplankton are a meroplanktonic group, exclusively dependent on the water column conditions and therefore their speciation events are mainly linked to a transgressive trend and to the CO_2 balance in the atmosphere/oceans system (Bartolini *et al.*, 1996). However, cyst producing dinoflagellates have a benthonic stage in their life cycle and are, therefore, influenced by the sea bottom conditions. Widespread anoxic conditions during a sea level rise produce a decrease in the available niches and favoured extinction events. The regressions may be related to dinoflagellate speciations if they are concomitant with the oxygenation of the sea bottom.

The high number of phytoplankton events recorded in central Italy makes possible a detailed correlation of the studied sections. In Fig. 2 the correlation lines based on integrated calcareous nannofossil and dinoflagellate cyst biostratigraphies are displayed. They reveal a variable accumulation rate among the different profiles and along the same succession. The accumulation rate is relatively high in the northern sector of the Umbria–Marche basin (Colle d’Orlando and Monte Serrone) with respect to the southern one (Somma and Fonte Cerro). The Pozzale section seems to show the most constant accumulation rate. A minimum thickness between the FO of *C. superbus* and *L. velatus* has been recorded in the expanded Monte Serrone succession. All the events inbetween have not been recorded. This may be the effect of a variation in the accommodation space, which prevented a large accumulation, or the effect of synsedimentary gravitational processes.

The biostratigraphical framework obtained in the present work yielded a detail greater than that provided by ammonites and generally independent from the sedimentary facies. Moreover, the integration of phytoplankton events may give preliminary information about the palaeoenvironmental evolu-

tion of a sedimentary basin, in spite of the scarcity of sedimentary structures and the apparent uniformity of vertical succession of lithotypes.

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Appendix A

Species index

Calcareous nannofossils

- Biscutum dubium* (Noël 1965) Grün in Grün *et al.* 1974
- Biscutum finchii* (Crux 1984) Bown 1987
- Biscutum grande* Bown 1987
- Biscutum novum* (Goy 1979) Bown 1987
- Bussonius prinsii* (Noel 1973) Goy 1979
- Bussonius lefuensis* Bown & Kielbowicz 1987 in Bown 1987
- Calyculus* sp. indet.
- Carinolithus poulnabronei* Mattioli 1996
- Carinolithus superbus* (Deflandre 1954) Prins in Grün *et al.* 1974
- Crepidolithus cavus* Rood, Hay & Barnard 1973
- Crepidolithus crassus* (Deflandre 1954) Noël 1965
- Crepidolithus granulatus* Bown 1987
- Discorhabdus criotus* Bown 1987
- Discorhabdus ignotus* (Gorka 1957) Perch-Nielsen 1968
- Discorhabdus striatus* Moshkovitz & Ehrlich 1976
- Lotharingius barozii* Noël 1973
- Lotharingius crucicentralis* (Medd 1971) Grün & Zweili 1980
- Lotharingius frodoi* Mattioli 1996
- Lotharingius hauffii* Grün & Zweili in Grün *et al.* 1974
- Lotharingius sigillatus* (Stradner 1961) Prins in Grün *et al.* 1974
- Lotharingius umbriensis* Mattioli 1996
- Lotharingius velatus* Bown & Cooper 1989
- Mitrolithus elegans* Deflandre 1954
- Mitrolithus jansae* (Wiegand 1984) Bown & Young in Young *et al.* 1986
- Mitrolithus lenticularis* Bown 1987
- Parhabdolithus liasicus* Deflandre 1952
- Schizosphaerella* sp. indet.
- Similiscutum crucilulum* De Kaenel & Bergen 1993
- Sollasites lowei* (Bukry, 1969) Rood *et al.* 1971
- Tubirhabdus patulus* Rood *et al.* 1973
- Watznaueria colacicchii* Mattioli 1996
- Watznaueria* sp.1 Cobianchi *et al.* 1992
- Dinoflagellate cysts
- Jansonia* sp.
- Luehndea spinosa* Morgenroth 1970
- Mencodinium semitabulatum* Morgenroth 1970 emend. Below 1987
- Mendicodinium brunneum* Bucefalo Palliani *et al.* 1997

- Mendicodinium microscabratum* Bucefalo Palliani *et al.* 1997
- Mendicodinium spinosum* subsp. *perforatum* Bucefalo Palliani *et al.* 1997
- Mendicodinium spinosum* subsp. *spiniosum* Bucefalo Palliani *et al.* 1997
- Mendicodinium umbriense* Bucefalo Palliani *et al.* 1997
- Ovalicysta cf. hiata* Bjaerke 1980
- Susadinium scrofoides* Dörhöfer & Davies 1980 emend. Below 1987
- Umbriadinium mediterraneense* Bucefalo Palliani & Riding 1997
- Valvaeodinium hirsutum* Bucefalo Palliani & Riding 1997
- Valvaeodinium koesseni* Morbey 1975 comb. nov. Below 1987
- Valvaeodinium punctatum* Wille & Gocht 1979 emend. Below 1987
- Valvaeodinium* cf. *perpunctatum* Wille & Gocht 1979 emend. Below 1987
- Valvaeodinium stipulatum* Wille & Gocht 1979 emend. Below 1987

Appendix B

Stratigraphical distribution of calcareous nannofossils in central Italy. Relative abundances have been calculated after a count of 250–300 specimens per smear slide. The number of views in which the standard quantity of specimens has been reached is taken into account for the total abundance evaluation.

Total abundance per field of view
V A = very abundant, more than 15 specimens
A = abundant, 10 to 15 specimens
C = common, 1 to 10 specimens
F = few, 0.1 to 1 specimens
R = rare, less than 0.1 specimens
B = barren
Relative abundance
V A = very abundant, more than 5 specimens in one field of view
A = abundant, 1 to 5 specimens in one field of view
C = common, 1 specimen in 1 to 10 fields of view
F = few, 1 specimen in 10–30 fields of view
R = rare, 1 specimen in more than 30 fields of view
X = 1 specimen in the smear slide
? = questionable presence
Preservation
G = good, without dissolution nor overgrowth effects
M = moderate, only minor effects of dissolution and overgrowth
P = poor, clear traces of dissolution/overgrowth
VP = diagenesis makes difficult the specific determination
Species richness
H = high, more than 15 species in the assemblage
M = medium, 5 to 15 species in the assemblage
L = low, less than 5 species in the assemblage

Fig. B1. Parameters used for the total and relative abundance evaluation; preservation and species richness classes.

Samples	Colle d'Orlando 1											Zone	
	Species	Schizosphaerella spp.											
		Total abundance	P. laeticus	T. paululus	M. lancea	M. elegans	M. leniticularis	C. crassus	C. covas	C. granulatus	B. dubium	B. planum	
14.40	VA VP H	CA . . . AR . R . . RR R . .									ARR . . R F R . . R . . .		
14.35	R P L	R	R										
14.10	R P L	R . . . R		R							R . . . R . . .		
13.95	R P L	R . . . R			R								
13.75	B												
13.50	R P L	R				R			R		R . . . R . . .		
13.30	R P L	R . . . R		R						R			
13.10	VA M H	VA . . A . R R R . R R R R F .					RCR . . .			RCRR . . RR			
12.90	VAP H	A . . A . R R . R R R R .						CRR . .		RCR . RRR			
12.70	A P H	CA . . A . R . R R R R .						CRR . .		RCRR . RR			
12.50	C P M C	R C R R R R R .							FR . .	RCR . . .			
12.30	C P H F	. C . . R R . R R R R .							CRR . .	RCRR RRR			
12.10	VAP M A	. A . . A . R R . R R .						CRR . .	RCRR . .				
11.90	VAP M VA	. A . . A . R R . R R R R .							RR . . . C . . .				
11.70	FC P M R	. F . . R . R R R R .							RR . . .	RC . R . . .			
11.50	A M H CA	. F . . R R . R R R R .							ARR . .	R F R R R . R			
11.30	C M H CAR	. R R . R R . R R R R .							CRR . .	R R R . . .			
11.10	C P H CA	. R R . R R . R R R R .							FRR R . . R R R R .				
10.90	A P H CA	. C . . R R R R R R .							CRR . .	R R R R . . .			
10.70	A M H CA	. R C . . R . R R R F F .							CRR . .	R R R R R . .			
10.50	A M H CA	. C . . R R F R F .								R C R R R . . .			
10.30	RF P M F	. R . . R . R R R R .							F R . . .	R R			
10.10	R PML	F . . R . . R . . R . . .											
9.70	VAPM H R	R R . R . R . R . R F F F .							RAFF R .	FC R R R R .			
9.50	FC P M FC	F C . . . R R R R R .							CRR . .	R R R R R . .			
9.30	C P H C	. F . R R . R R R F R .							CF RR .	R F R . . . RR			
9.10	FC P M C	. F . R . R . R R R R .							RRR . .	R R			
8.90	VAG H C	. F . R . R . R R F F F R A F F R .								R F R R . . .			
8.70	F M H F R	. R . R R R R R R R R C F R R .								R R R			
8.50	VAP H R	. R . . R . F C F R F .							AF RR .	RC RR . . R			
8.30	C M H R	. R F . R . . R R F R R R R R A F F R .								F F R R . . RR			
8.10	C P M C	. F . . R R R R R .							CR FR . .	R R			
7.90	VAPM H C	R . A . R R . R R F R F .							RA F F . .	R F R R . . .			
7.70	VAP H A	. A . A . R . R R F F .							RA F R R R R F R R .	R R			
7.50	A P H FC	R . F . R . R . R R R ? R A F R R .							R F R R . . .				
7.30	VAPM H C	R . A . R . F F F F F R F A F F R .								RC F R . . .			
7.10	VAP H R	R R . . F R C R C .							RA F R R . .	R F R R F . . R			
6.90	VAPM H A	R . A . R . R R R F .							RA F R R . .	FC F . R . . .			
6.70	VAPM VH A	R R C . R R R R R R R R ? R C R R R .								RC RR			
6.50	F M M F	F . . R . R R R R R .								F R R R R R R R .	R		
6.30	A P M A	. F . . R R R R R .								RC R . . . R .	RR		
6.10	VAPM H VA	. F . R R . R R R R R R C R R R .								R F R . . . R .	R R		
1.90	F VP M C	. F . F . R . R . . F . .									F R . . . F . . .		
1.70	F P M C	. R . F . F . . R . . F R R .									RC . . . R . . .		
1.60	F PM M CA	. R . F . R . F . . C R R . . RR . . . C .											
1.50	F PM M CA	R . F . R . F . . C R R . . RR . . . C .											
1.40	RF P L C	. F . C R . . .									R		
1.20	RF VP M C	. F . F . . F . . F R . . .									R C R		
1.10	C PM M A	. F . C F . . F R . A . . R F F .											
1.00	RF P M A	. F C . F R . . R R . . F R . R . C R . . F .											
0.90	F P M A	. C . C . R . F . . CF . . RF . . .											
0.80	F P M CA	. C . C . C . F . . R . . CF . . RF . . .											
0.75	C P M A	R . A R R R . . R R R R . . ARR R . R . . .											
0.70	F P M A	. F C . F . . R R . . RR . . CF . F . F F R . . .											
0.60	RF P M C	. A . C . R R F . . A . R . . F R . . .											
0.50	C P M C	R A R R . R . R . R . R . C											
0.40	C P M CA	A . A . R . R . R . R . R . R . C . . R R . . .											
0.30	C P M CA	R R . R . R . R . R . R . R . A R R . R . R . . .											
0.20	FC P M A	R . A . R R . R . R . R R . C . . . R . . .											
0.10	FC P M A	R A R R . R . R . R R R . C . . . F . . .											
0.00	C M M A	R R A . R R . R R . R R . C . R . R . . .											

D. tenunicostatum

Fig. B2. Calcareous nannofossil distribution in the Colle d'Orlando section, lower part.

Colle d'Orlando 2								Zones	
Samples	Species	Total abundance	Preservation	Species richness					
				Schizospiraerella spp.					
				<i>P. Nasuta</i>	<i>T. penaius</i>	<i>M. tiansae</i>	<i>M. elegans</i>		
41.50	C	P	M	A		X			
41.00	F	VP	L	C		R	.		
37.65	R	VP	L	C		R	.		
37.60	FC	P	M	F	R	.			
35.30	A	PM	H	A	FF	CF	C		
33.30	F	M	PM	F	.		RR		
31.10	F	P	M	C	.	R	X		
30.30	F	M	P	F	.	X	X		
30.05	F	M	M	F	.	X	X		
29.60	F	M	M	F	.	FR	FR		
28.30	C	M	H	C	.	FR	F		
27.30	F	P	M	F	.	FF	FR		
26.50	F	P	M	F	.	RR	FF		
25.60	FC	P	H	F	.	RXX	FRRX		
25.10	F	P	H	F	.	RXX	FRRX		
24.55	F	P	M	F	.	RXRR	XR		
24.00	R	P	M	R	.	R	X		
23.60	F	PM	M	F	.	X	X		
22.95	C	G	H	C	.	FRX	XRF		
22.90	RF	P	L	F	.	.	.		
22.70	RF	P	L	F	.	.	R		
22.30	VAP	H	VAR	.	RRR	F	R		
21.90	A	PM	H	VAR	RR	F	RRF		
21.70	CA	P	H	A	.	RRRR	RFR		
21.50	FC	P	M	CA	.	RRR	R		
21.30	FC	P	M	CA	.	F	RR		
21.10	A	P	H	C	R	R	FRR		
20.90	F	VP	M	R	RRR	RFR	ARR		
20.60	VR	L	VR	.	.	.	R		
20.50	C	P	M	A	.	F	RRR		
20.30	R	P	L	F	RR	.	F		
19.90	FC	VP	M	C	.	FR	RR		
19.70	A	P	H	R	RRR	F	FFFF		
19.50	A	P	H	VA	.	RR	CFRR		
19.30	A	PM	M	VA	.	F	RR		
19.10	A	P	M	VA	.	R	RR		
18.90	A	P	M	A	.	FR	RR		
18.70	B		
18.30	FC	P	M	C	.	RRRR	RR		
18.10	R	VP	M	R	.	RR	R		
17.90	C	P	M	CA	RRRR	RR	RR		
17.70	VR	L	VR		
17.50	B		
17.30	VR	L	VR		
17.10	VR	L	VR	.	.	.	R		
16.70	R	P	M	R	RR	R	F	R	
16.50	B		
16.30	FC	M	M	FC	.	A	RR		
16.10	C	MG	M	C	R	A	RR		
15.90	FC	M	M	C	.	A	RR		
15.70	B	R	R	
15.60	B	R	R	
15.40	VR	L	R		
15.20	VR	L	R		
15.00	VR	L	R		
14.80	C	VP	M	FC	.	ARR	R		
14.75	FC	VP	M	FC	.	C	RRRR		
14.60	CA	VP	H	FC	.	ARR	RRRRR		

Fig. B3. Calcareous nannofossil distribution in the Colle d'Orlando section, upper part.

*D. tenuecostataum**H. serpentinus*

Monte Serrone 1					
Samples		Species		Schizosphaerella spp.	
	Total abundance		Species richness		Preservation
15.30	V A	H P	V A R . R . R	<i>P. llaisticus</i>	
17.50	V A	M PM	V A R . R . R R	<i>M. jansae</i>	
18.00	V A	M P	V A R . R . R R	<i>M. elegans</i>	
18.30	V A	M VP	V A . R . F	<i>M. lenicularis</i>	
18.80	V A	H P	V A R F R . R R .	<i>T. pantulus</i>	
18.95	V A	M PM	V A R	<i>C. crassus</i>	
19.02	V A	H P	V A R R F	<i>C. canus</i>	
19.20	V A	H MG	V A R R R	<i>C. granulatus</i>	
19.40	V A	M VP	V A R R	<i>B. dubium</i>	
19.60	V A	M P	V A F	<i>B. novum</i>	
19.65	V A	M VP	V A R R R	<i>B. finchii</i>	
19.90	V A	H P	V A R . R F R R .	<i>B. grande</i>	
20.55	V A	M VP	V A R R	<i>S. lowei</i>	
20.80	V A	M P	V A R R R	<i>L. hauffii</i>	
21.37	V A	H PM	V A R F	<i>L. umbrienensis</i>	
21.95	V A	H P	V A R R	<i>B. prinsii</i>	
22.50	A M	P A	R R R	<i>L. frondo</i>	
22.70	A M	PM A	R R R	<i>L. barozzi</i>	
23.25	C M	P A	A R R	<i>Calyculus</i> spp.	
23.75	A M	PM A	A R	<i>C. sigillatus</i>	
25.20	V A	M M	V A R F R R R R .	<i>L. crucicentralis</i>	
25.50	F M	P C	R . R	<i>B. leijenensis</i>	
26.80	C M	P A	R R R . R	<i>C. poulnabronei</i>	
27.85	A M	P A	F . R . R R	<i>C. superbus</i>	
					<i>D. velutinus</i>
					<i>D. ignotus</i>

Fig. B4. Calcareous nannofossil distribution in the Monte Serrone section.

Monte Serrone 0					
Samples		Species		Schizosphaerella spp.	
	Total abundance		Species richness		Preservation
4.90	V A	H M	V A R R R . R R R	<i>P. llaisticus</i>	
3.70	V A	M P	V A R	<i>M. jansae</i>	
3.35	V A	M PM	V A R	<i>M. elegans</i>	
2.75	V A	H M	V A R R R . R F R R R R .	<i>M. lenicularis</i>	
2.30	V A	H PM	V A R R R R R F .	<i>T. pantulus</i>	
2.15	A M	P V A	R . R R . R	<i>C. crassus</i>	
1.80	V A	H P	V A R R . R R F .	<i>C. canus</i>	
1.65	V A	M PM	V A R R . R R F .	<i>B. dubium</i>	
1.30	V A	H P	V A R R . R F . R R F R .	<i>B. novum</i>	
0.65	V A	H P	V A R . R F . R F . R R F .	<i>B. finchii</i>	
0.35	A M	VP V A	F . R . F . R F . R R F .	<i>S. lowei</i>	
0.20	V A	H PM V A	R F . R . R R A R F R R R R R .	<i>L. hauffii</i>	
				<i>L. umbrienensis</i>	
				<i>L. frondo</i>	
				<i>L. barozzi</i>	
				<i>Calyculus</i> spp.	
				<i>C. sigillatus</i>	
				<i>L. crucicentralis</i>	
				<i>B. prinsii</i>	
				<i>B. leijenensis</i>	
				<i>C. poulnabronei</i>	
				<i>C. superbus</i>	
				<i>L. velutinus</i>	

Fig. B5. Calcareous nannofossil distribution in the Monte Serrone section, southern sector.

Samples	Pozzale						Zones
	Species	Total abundance	Preservation	Species richness			
		C	P	M	L	A	
10.40	FC	P	M	CA	.	R.	
10.15	F	VP	L	C	.	R.	
9.55	C	VP	M	C	.	CR.	
9.15	A	P	H	.	RRR.	RRRR.	
8.90	VAP	FM	M	VA	.	R.	
8.50	CA	P	M	A	.	R.	
7.70	A	P	M	VA	.	R.	
7.35	CA	P	M	A	.	R.	
7.30	VA	M	H	VA	.	R.	
7.20	VA	MGM	VA	.	F.	CR.	
7.10	A	PM	M	AR	.	RFR.	
7.00	A	PM	M	A	.	RFR.	
6.90	CA	P	M	CA	.	R.	
6.80	FC	P	L	A	.		
6.70	VA	M	H	VA	.	F.	
6.60	C	P	M	CA	.	F.	
6.50	A	P	M	CAR	.	FR.	
6.40	A	P	M	A	.	F.	
6.20	C	P	M	CA	.	R.	
6.10	C	P	M	A	.	R.	
6.00	VR	VP	L	R	.		
5.90	VR	VP	L	R	.		
5.80	CA	P	M	A	.	R.	
5.70	R	VP	L	F	.	R.	
5.60	A	P	M	A	.	R.	
5.50	A	P	M	A	.	R.	
5.40	A	PM	M	VA	.	C.	
5.30	F	PM	C	CR	.	R.	
5.20	R	P	M	F	.	RRR.	
5.10	C	PM	H	CR	RRR.	RCRRRRRRRR.	
5.00	A	P	M	A	.	RFR.	
4.90	F	P	L	CA	.	F.	
4.80	F	P	L	CA	.	R.	
4.70	F	P	M	CA	.	R.	
4.60	F	P	M	CA	.	R.	
4.50	A	P	M	VA	.	C.	
4.40	A	P	M	A	.	C.	
4.30	A	M	M	A	.	A.	
4.20	A	P	M	A	.	C.	
4.10	C	P	L	C	.	C.	
4.00	A	PM	M	A	.	A.	
3.80	A	P	M	A	.	F.	
3.70	VA	M	H	VA	.	A.	
3.60	A	P	M	VA	.	C.	
3.50	VA	PM	H	VAR	.	R.	
3.40	A	VP	M	VA	.	A.	
3.30	VAP	M	VA	.	A.	R.	
3.20	A	VP	M	VA	.	R.	
3.10	VAP	P	H	VA	.	C.	
3.00	A	PM	H	VA	.	A.	
2.90	VAP	M	VA	.	A.	R.	
2.80	VAP	PM	VA	.	A.	R.	
2.70	VAP	PM	VA	.	A.	R.	
2.60	A	P	H	VA	.	C.	
2.50	R	P	L	C	.	R.	
2.24	C	P	M	A	.	R.	
2.00	VR	VP	L	R	.		
1.40	C	P	L	A	.	R.	
1.30	C	P	L	C	.	R.	
0.90	R	P	L	F	.	R.	
0.85	R	P	M	C	.	R.	
0.60	R	P	L	C	.	R.	
0.50	R	P	L	F	.	R.	
0.40	R	P	L	C	.	R.	
0.30	R	P	M	C	.	R.	
0.00	R	R	L	C	.	R.	

Fig. B6. Calcareous nannofossil distribution in the Pozzale section.

Samples	Species			Somma																								Substage							
				Total abundance			Species richness																												
				Preservation																															
2.90	VA	H	M	VA	.	.	<i>Schizospherella</i> spp.																												
2.85	VA	H	M	VA	X	.	<i>P. hispanicus</i>	.	.																										
2.70	VA	H	PM	A	R	R	<i>T. patulus</i>	.	.																										
2.60	VA	H	MG	A	.	R	<i>M. jansae</i>	.	.																										
2.55	A	M	PM	VA	.	.	<i>M. elegans</i>	.	.																										
2.40	C	M	PM	CA	.	.	<i>M. lenticularis</i>	.	.																										
2.35	A	M	VP	A	.	X	<i>C. cavaus</i>	.	.																										
2.30	VA	H	PM	A	.	X	<i>B. dubium</i>	.	.																										
2.20	A	H	M	F	.	XX	<i>B. finchii</i>	.	.																										
2.05	VA	H	PM	A	R	R	<i>B. novum</i>	.	.																										
2.00	A	M	P	R	.	.	<i>S. cracium</i>	.	.																										
1.95	VA	H	P	R	R	R	<i>B. grande</i>	.	.																										
1.85	A	H	VP	FC	R	X	<i>S. orbicularis</i>	.	.																										
1.78	VA	H	PM	VA	R	R	<i>S. lowei</i>	.	.																										
1.75	C	H	P	R	.	X	<i>L. hauffii</i>	.	.																										
1.71	A	H	P	VA	.	X	<i>L. umbriensis</i>	.	.																										
1.60	A	M	P	VA	.	X	<i>L. frodoi</i>	.	.																										
1.52	A	M	PM	VA	.	R	<i>L. cruciferatris</i>	.	.																										
1.35	VA	M	M	VA	X	X	<i>B. prinsii</i>	.	.																										
1.15	A	M	PM	VA	.	F	<i>B. lequensis</i>	.	.																										
.85	A	M	PM	VA	.	F	<i>C. polynathonei</i>	.	.																										
.65	A	M	PM	VA	X	R	<i>C. superbas</i>	.	.																										
.55	A	M	VP	A	.	R	<i>L. velatus</i>	.	X	.																									
.40	A	L	PM	A	.	R	<i>D. ignorans</i>	.	.																										
.10	A	M	VP	VA	.	R	<i>D. constans</i>	.	X	.																									
.01	A	M	PM	VA	R	R	<i>R. XX</i>	.	.																										

Fig. B7. Calcareous nannofossil distribution in the Somma section.

Fonte Cerro'							
Samples		Species		Species richness			
Total abundance	Preservation	Species richness		Schizophaerella spp.	<i>P. hastatus</i>	<i>T. parvulus</i>	<i>M. japonica</i>
5.55	R P	L	R
5.40	VA PM H	VA	.	F	F F C R R A	R R R F	.
5.35	VA PM M	VA	.	F	R R R R	C R	.
5.25	VA PM M	VA	R	F	R R F	A R R	.
5.15	R P L	F	.				
5.00	VA M M	VA	R	F	R R R	C R R	.
4.90	A P M A	R	.	F	R R R	R C R R	.
4.80	C P M A	R	.	R	R	R R	.
4.70	C P M A	.	.	R	R R	R R R	.
4.60	VA P M	VA	R	R	R R	R F	.
4.40	A P L	VA	.	R	R	F R	R
4.30	VA P M	VA	R	F	R F	R R	.
4.20	VA PM M	VAR	.	R F	R R	C R	.
4.15	VA M M	VA	R	R F	.	C R R	R
4.00	CA P L A	.	.	R	.	R R	.
3.90	CA P M A	R	.	R	.	R R R	R
3.85	CA P L A	F	R
3.55	A VP M A	R	.	R	R	F R	.
3.45	VA P H	VAR R R	.	R R	F R R F R	A R R	R F R
3.40	A P M VA	.	.	R R R	R	F R	.
3.30	A P M VA	R	.	R	R R	R R R	.
3.15	VA P M VA	.	.	R	R R	C R R	.
3.00	VA PM M	VA	R	C	R R R	A R R	R
2.90	VA M H	VA	R	C R	R F R	A R R R R R	R R R
2.80	A P M VA	.	.	R R	R R	C R R R R	R
2.70	VA PM M	VA	.	F R	R R R	C R R R R R	R
2.65	VA PM M	VA	R	F	R	C R R	R R R
2.55	VA M M VA	.	.	F	F	R F R	C R R
2.40	VA PM M	VA	.	F	R	F R R	R
2.30	VA PM M	VA	.	F R	R R	F R R	R R
2.15	A P L VA	.	.	R	.	R	.
2.05	A P M VA	R	.	R	R	R R	R R
1.90	VA P M VA	.	F	.	R R R	R R	R
1.80	VA P M VA	.	F	.	R	F R	R
1.70	A P L VA	.	F	.	F	.	.
1.55	A P M VA	R	F	R R	F R	.	R
1.45	VA M M VA	.	C R	R R	C R R	R R R	R R R
1.35	VA PM M VA	.	F	.	R R R	R R	.
1.25	VA P M VA	.	C R R R	R R R	F R R	R R	R
1.20	VA P M VA	R	R C R	.	F	.	.
1.10	A P L A	.	R F R	.	.	R	.
0.70	C P L A	C	R	.	R	.	.
0.60	VA PM M A	A	R	R R	C R R R R F R R	.	?
0.50	A P L A	C	R	R	R	.	.
0.40	VA P M VA	C	R R	R R R	F R	F	R
0.35	VA P M A	C	R R	R R	R F R	C	R R
0.20	F VP L C
0.10	F VP L C	C	R R	.	R	R	.
0.00	C P M A	F	.	R R	R R	R R	R ?
H. serpenitus							
D. tenuicostatum							
Fonte Cerro							
2.60	VAP L VA	.	F	R	.	R	.
2.45	VAP M VA	R R	R R	.	R R	.	R
2.30	VAM H VA	R R R R R	R R R R R	F R R R R R	.	.	?
2.20	VAP M H VA	R F	F R R F R F R R	C R R R R R	?	R R	.
2.10	VAP M VA	R F	R F F	F R F F	C R R	R	R
2.00	VAP H VAR R R	R	R R R R R	C R R R R	?	.	.
1.90	A P M VAR R R	R R	R	R R	R R	.	.
1.75	A P M VAR R R	R	R R	R R	F R	.	.
1.48	A VP M VA	F	R	R R	R R R	?	.
1.35	FC VP M A	R	R	R R	R	.	.
1.20	C VP L A	R	R R	.	R	.	.
0.90	A VP M VA	R F	.	R R	R	.	.
0.80	VAP M VA	R F	R R R	R R	F R R	.	.
0.60	A VP M VA	R R	R	R	R R	?	.
0.45	C P M A	R F	R	R R	F R R R	.	.
0.40	C VP L A	R R	.	R	R	.	.
0.30	FC VP M A	R R	R R	R	R	.	.
Lower Toarcian							
Upper Pliensbachian							

Fig. B8. Calcareous nannofossil distribution in the Fonte Cerro section.

Appendix C

Semi-quantitative distribution of dinoflagellate cysts in the Lower Jurassic of central Italy. The abundances have been estimated by averaging the number of dinoflagellate cysts counted for each species in three slides. A species was classified rare (R) if it averaged between 0 and 5, common (C) if it averaged between 5 and 10, frequent (F) if it averaged between 10–20 and abundant if it averaged more than 20 dinoflagellate cysts.

COLLE D'ORLANDO section		
LOWER JURASSIC	SUBSYSTEM	SUBSTAGE
	Ammonite Zone	
	Samples	
	Species	
	<i>M. spinosum</i> subsp. <i>spinosum</i>	
	<i>Mendicodium microscabratum</i>	
	<i>Mendicodium umbriense</i>	
	<i>Valvocodium cf. stipulatum</i>	
	<i>Jansonia exigua</i>	
	<i>Valvocodium hirsutum</i>	
	<i>Umbridiadnum mediterraneense</i>	
	<i>Valvocodium jurasicum</i>	
	<i>Polygonium punctatum</i>	
	<i>Valvocodium brunnneum</i>	
	<i>Mancodium semitubulatum</i>	
	<i>M. spinosum</i> subsp.	
	<i>Valvocodium sp.</i>	
	<i>Susadinium acrofoides</i>	
24.70		R
23.50		R
21.90		R
19.30		C
18.90		R
18.10		C
17.50		R
16.90		R
15.90		R
14.80		A
		A
		C
13.10		R
		R
		R
12.70		C
		R
		R
12.10		R
11.70		C
		R
		R
10.50		R
		R
9.10		R
8.50		C
		R
7.70		
7.30		R
6.90		R
6.10		R
		A
		R
		R

Fig. C1. Dinoflagellate cyst distribution in the Colle d'Orlando section.

MONTE SERRONE section		
LOWER JURASSIC	SUBSYSTEM	SUBSTAGE
<i>H. serp.</i>	Ammonite Zones	Species
<i>D. tenuicostatum</i>	MS1-1.00	<i>Umbridiadnum mediterraneense</i>
	MS0 8.50	<i>Valvocodium hirsutum</i>
	MS0 8.00	<i>Mendicodium microscabratum</i>
	MS0 7.00	<i>Mendicodium umbriense</i>
	MS0 1.20	<i>Valvocodium sp.</i>
		<i>Mendicodium microscabratum</i>
		<i>Mendicodium brunnneum</i>
		<i>Mendicodium semitubulatum</i>
		<i>M. spinosum</i> subsp. <i>spinosum</i>

Fig. C2. Dinoflagellate cyst distribution in the Monte Serrone section.

POZZALE Section		
LOWER JURASSIC	SUBSYSTEM	SUBSTAGE
<i>H. serp.</i>	Ammonite Zones	Species
<i>D. tenuicostatum</i>	7.00	<i>Mendicodium microscabratum</i>
	5.70	<i>M. spinosum</i> subsp. <i>spinosum</i>
	4.00	<i>M. spinosum</i> subsp. <i>microreticulatum</i>
	3.60	<i>Mendicodium umbriense</i>
	3.50	<i>Umbridiadnum mediterraneense</i>
	3.40	<i>Valvocodium hirsutum</i>
	3.10	<i>Luehinea spinosa</i>
	3.00	<i>Polygonium jurasicum</i>
	2.80	<i>Mancodium semitubulatum</i>
	2.60	<i>Valvocodium punctatum</i>
		<i>Mendicodium brunnneum</i>
		<i>Susadinium scrobifides</i>
		? <i>Ovalicysta</i> cf. <i>hirtata</i>

Fig. C3. Dinoflagellate cyst distribution in the Pozzale section.

SOMMA section						
		SUBSYSTEM		Species		
		SUBSTAGE		Samples		
LOWER JURASSIC						
	Lower Toarcian	MBE 2.20		<i>Mendicodinium bruneum</i>	A A	
		MBE 2.05		<i>Polygonum jurasicum</i>	C	
				<i>Mendicodinium microscabrum</i>	R R	
				<i>Mendicodinium umbriense</i>		
				<i>Valvaeodinium hirsutum</i>		
				<i>Umbriadinium mediterraneense</i>		

Fig. C4. Dinoflagellate cyst distribution in the Somma section.

FONTE CERRO section						
		SUBSYSTEM		Species		
		SUBSTAGE		Ammonite Zones		
LOWER JURASSIC				Samples		
Late Pliens.	Lower Toarcian	<i>D. tenuicostatum</i>		<i>Mendicodinium microscabrum</i>		
		<i>P. spin.</i>	FC" 2.70	<i>M. spinosum</i> subsp. <i>spinosum</i>		
			FC" 2.05	<i>M. spinosum</i> subsp. <i>microreticulatum</i>		
			FC" 1.40	<i>Mendicodinium umbriense</i>	R	
			FC" 1.20	<i>Valvaeodinium koessensi</i>	R R	R R
			FC" 1.00	<i>Mancodinium semitubulatum</i>	R R R	R R R
			FC" 0.85	<i>Lithedida spissa</i>	R	
			FC" 0.70	<i>Valvaeodinium hirsutum</i>	R R C C R	
			FC" 0.40	<i>Umbriadinium mediterraneense</i>	R	
			FC" 0.15	<i>Polygonum jurasicum</i>	R	
			FC" 2.10	<i>Mendicodinium bruneum</i>		
			FC" 0.40	<i>Valvaeodinium punctatum</i>		
			FC" 0.25	<i>Sasadirnum scrofoides</i>		

Fig. C5. Dinoflagellate cyst distribution in the Fonte Cerro section.

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